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A point mutation in the AF-2 domain of thyroid hormone receptor alpha1 expressed after CRE mediated recombination partially recapitulates hypothyroidism.

Short title: TR α 1 dominant negative mutation in mice.

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Abbreviations footnote: Q-RT-PCR: quantitative-reverse transcription-polymerase chain reaction.

ABSTRACT

Thyroid hormones act directly on transcription by binding to TR α 1, TR β 1, TR β 2 nuclear receptors, regulating many aspects of post-natal development and homeostasis. To precisely analyze the implication of the widely expressed TR α 1 isoform in this pleiotropic action, we have generated transgenic mice with a point mutation in the TR α 1 coding sequence, which is expressed only after CRE/loxP mediated DNA recombination. The amino-acid change prevents interaction between TR α 1 and histone acetyltransferase coactivators and the release of corepressors. Early expression of this dominant-negative receptor deeply affects post-natal development and adult homeostasis, recapitulating many aspects of congenital and adult hypothyroidism, except in tissues and cells where TR β 1 and TR β 2 are predominantly expressed. Both respective abundance and intrinsic properties of TR α 1 and TR β 1/2 seems to govern specificity of action.

INTRODUCTION

Thyroid hormones (TH) consist in 3,5,3'-tri-iodothyroinine (T3) and its precursor thyroxine (T4) which displays a weaker biological activity. Both molecules are secreted by thyroid gland follicles, but most of T3 synthesis results from T4 deiodination in other organs. T3 binds to nuclear receptor (TRs) present as three different isoforms, TR α 1, TR β 1, TR β 2 encoded by the two *THRA* and *THRB* genes. TRs bind in a ligand-independent manner to specific response elements, mainly as RXR heterodimers, which are widespread in the genome. Unliganded TRs recruit transcription corepressors. Among these NcoR and SMRT function as platforms for the recruitment of histone deacetylases. T3 binding results in a displacement of the C-terminal helix of TR, which contains activation function 2 (AF-2). This permits the recruitment of several coactivators complexes, some of them, like SRC1 and SRC2, having a histone acetyl-transferase activity (1, 2) and destabilizes interactions with histone deacetylase corepressors.

TH signaling exerts multiple effects on post-natal development and the maintenance of homeostasis in adults by directly regulating target gene transcription (3). Previous investigations used various strategies to decrease TH levels. Among several animal models, transgenic mice recently gained popularity, at the expense of thyroidectomized or pharmacologically manipulated rats. In *Pax8*^{-/-} knockout mice, the only reported primary defect is the absence of thyroid follicular cells (4). Although these animals can receive maternal TH through placenta during fetal development, they usually die within three weeks after birth, unless they are rescued by TH treatment. The direct and indirect effects of congenital hypothyroidism are difficult to unravel. TR knockout mice offered new possibilities to solve this problem and allowed attributing specific function to each receptor isoform

(5). The broadly expressed TR α 1 appears to be the main regulator of development during the first three weeks of post-natal, pre-weaning development. This period, somewhat reminiscent of amphibian metamorphosis (6), is marked by a transient increase in TH circulating level. At this time, liganded TR α 1 regulates intestinal remodeling (7), cerebellum development (8), spleen erythropoiesis (9) and bone growth (10). TR α 1 has a later role in setting cardiac function (11) and thermogenesis. TR β 1 and TR β 2 expression pattern is more restricted. These isoforms are the main regulators of liver function, inner ear development, retinal cones differentiation, and feedback regulation of the hypothalamic-pituitary-thyroid axis.

Surprisingly the deletion of all TR isoforms, obtained by the combination of *THRA* and *THRB* knockouts, is not lethal and only partially recapitulates the post-natal consequences of congenital hypothyroidism observed in *Pax8*^{-/-} mice (12, 13). Several studies support the hypothesis that this discrepancy is due to the negative effect exerted by the unliganded TR α 1 present in hypothyroid *Pax8*^{-/-} mice, but not in *THRA/THRB* knockout mice. In line with this, the consequences of TH depletion are attenuated when *THRA* is deleted (8, 14). Furthermore the consequences of *THRA* knockout are limited compared to those of *THRA* knock-in mutations. Three germline knock-in mutations have been introduced in the ligand binding domain of TR α 1. The reading frame mutations have different consequences on ligand binding and cofactor interactions, but all preserve DNA binding and reduce the transcriptional activation ability of TR α 1. Mice heterozygous for these mutations display many features of congenital hypothyroidism, suggesting a constitutive repression of TH target genes (15-18). The hypothesis that congenital hypothyroidism is mainly due to the negative action of unliganded TR α 1 remains however controversial for several reasons. First, the rescue of *Pax8*^{-/-} mice by *THRA* knockout is only partial, and not observed with all *THRA* alleles (19). Another difficulty is that the consequences of the three *THRA* knock-in mutations are not identical. For example, obesity has been reported in only one case. For these reasons, several additional hypothesis have been proposed, including the intervention in hypothyroid mice of unliganded TR β (20), of TR α 2, a *THRA* encoded isoform which is unable to bind T3 (19), or of TH non-genomic effects (21, 22).

We describe here the construction of a new mouse allele for the *THRA* gene, encoding a TR α 1 receptor with an AF-2 mutation (L400R). According to structural data, the mutation fills a cavity at the surface of the ligand binding domain which is required for histone acetyl transferase coactivators interaction. The mutation prevents histone acetyl transferase recruitment and favours the permanent recruitment of corepressors. This is responsible for a dominant negative activity exerted on both TR α 1 and TR β receptors in transient expression assays. The genetic construct introduced by homologous recombination in the *THRA* locus possesses an upstream floxed cassette, allowing for a spatio-temporal expression control of TR α 1^{L400R} expression, using the Cre/loxP recombination system. The

phenotypic consequences of an early embryonic recombination of this *THRA* allele on development and homeostasis closely resemble those of congenital and adult hypothyroidism, except in tissues where *THRB* function is predominant. We conclude that, although hypothyroidism manifestations mainly results from the negative action of the ubiquitous unliganded TR α 1, unliganded TR β also contribute in several tissues.

RESULTS

Construction of the *TR α ^{AMI}* allele

We introduced in the AF-2 domain of the mouse TR α 1 receptor a point mutation converting a leucine into an arginine (L400R) equivalent to the L454R mutation previously created in the human TR β 1 (23, 24). It has been shown previously that this TR β 1 mutation fully prevents interaction with the histone acetyl transferase coactivators while preserving interaction with histone deacetylase corepressors (34), resulting in a very strong dominant-negative activity on TR mediated transactivation. Recently, this mutation has also been shown to prevent an interaction between AF-2 and the hinge region, but the functional significance of this interaction remains unclear (25). We verified, using a proteolysis sensitivity assay, that ligand binding was not abolished by the mutation (data not shown). We used yeast two-hybrid assays to confirm that, whether T3 is present or not, TR α 1^{L400R} interacts with the NcoR histone deacetylase corepressor but not with the SRC1 or SRC2 histone acetyl transferase coactivators (Figure 1A). We then introduced the TR α 1^{L400R} cDNA between two cassettes. The downstream *IRESTaulacZ* cassette encodes a β -galactosidase targeted to the cytoskeleton, translated from an ECMV internal ribosomal entry site. We placed the entire construct downstream to a CMV transcription promoter (figure 1B) to perform transient expression assays. The upstream *PGKNeoRpolyA* cassette confers G418 resistance to eukaryotic cells and contains a SV40 polyadenylation signal, which arrests most transcription and prevents any TR α 1^{L400R} translation. As the upstream *PGKNeoRpolyA* cassette is flanked by two tandem *loxP* sequences, CRE mediated recombination is required to produce a bicistronic mRNA encoding both TR α 1^{L400R} and β -galactosidase. This design was expected to favor the detection of recombination events in mouse tissues by Xgal staining. Transient expression results confirm that synthesis of both TR α 1^{L400R} protein (Figure 1C) and β -galactosidase (data not shown) requires previous excision of the *PGKNeoRpolyA* cassette. TR α 1^{L400R} was not able to activate transcription via a T3 response elements (2xDR4), and exerted a strong dominant negative effect on transcriptional activation mediated by the wild-type liganded TR α 1 receptor (Figure 1D). A similar dominant-negative action was observed when TR β 1 or TR β 2 was used for transactivation (data not shown). All these observations lead to the conclusion that the L400R mutation places the TR α 1 receptor in a conformation, equivalent to the unliganded state, that exert a constitutive dominant-negative effect on TR target genes expression.

The TR α 1^{L400R} reading frame flanked by the upstream *PGKNeoRpolyA* and the downstream *IRESTaulacZ* cassettes, was introduced in the mouse *THRA* locus by homologous recombination (Figure 2A,B). In the resulting *TRa^{AMI}* allele (AF-2 Mutation, Inducible), the mutation is silent. CRE recombination is required to express TR α 1^{L400R} from its natural promoter, at a level similar to the endogenous TR α 1 receptor. To address the ability of ES cells to express *TRa1^{L400R}* after CRE recombination, CRE was transiently expressed in *TRa^{AMI}/+* ES cells and RT-PCR was performed to amplify the 3' part of the TR α 1 cDNA. As *TRa1^{L400R}* cDNA differs from its wild-type counterpart by the presence of a SacII restriction site we digested the PCR product with SacII and confirmed by gel electrophoresis the presence of both type of mRNA in *TRa^{AMI}/+* ES cells after CRE recombination (Figure 2C). ES cells carrying an intact *TRa^{AMI}* allele were injected into blastocytes to produce mouse chimeras. After germline transmission, *TRa^{AMI}/+* mice, which express only one intact *THRA* allele, were obtained. They displayed, as expected, a normal growth and fertility and an apparently normal phenotype. *TRa^{AMI}/TRa^{AMI}* animals, which do not express any *THRA* isoforms, were obtained at the next generation. These homozygous animals, whose phenotype was not investigated, were able to develop and breed normally, like other *THRA* knockout mice.

Early and ubiquitous expression of TR α 1^{L400R} leads to a hypothyroid-like phenotype

To trigger a broad expression of TR α 1^{L400R} we crossed *TRa^{AMI}/+* mice with *SYCP1CRE* transgenic mice. These mice have been reported to express the CRE recombinase only during male spermatogenesis (26). We thus expected *TRa^{AMI}/+xSYCP1CRE* males to give rise to progenies where a deleted *TRa^{AMI}* allele would be in all cells from the beginning of development, and TR α 1^{L400R} expressed ubiquitously. Unlike previous reports, we found that complete excision did not occur in all cases, some *TRa^{AMI}/+xSYCP1CRE* males giving rise to offspring with no or partial excision. To circumvent this problem, probably due to variegation in CRE expression, we selected favorable *TRa^{AMI}/+xSYCP1CRE* males for further experiments and only considered pups with excision efficiency, as evaluated by PCR, superior to 90% (Figure 3A). We will call these heterozygous mice, which express *TRa1^{L400R}* in most tissues, *TRa^{AMI}xS*. We performed RT-PCR to demonstrate the presence of a SacII containing *TRa1^{L400R}* cDNA in several tissues (data not shown). Q-RT-PCR was also used to evaluate the total expression level of *TRa1* (*TRa1^{L400R}*+wild-type) in *TRa^{AMI}xS* mice. The expression level at P15 was similar to the one observed in wild type animal in liver with a 3-fold increase in the cerebellum (Table I) and brain (data not shown). This slight augmentation could result from the absence of alternate splicing in the *TRa^{AMI}* locus, a phenomenon that normally diverts part of the *THRA* transcription to produce TR α 2, an isoform which does not bind T3. β -galactosidase activity was very weak or undetectable, probably due to low expression level of the *THRA* locus, and perhaps to limited efficiency of translation initiation at the IRES element.

At P15 the fraction of $TR\alpha^{AM1}xS$ in litters was close to the expected 50% ratio. It seems therefore that expression of the $TR\alpha1^{L400R}$ expression is compatible with pre-natal and early post-natal development of heterozygous mice. By contrast later development was deeply affected, and 48% of the $TR\alpha^{AM1}xS$ pups did not reach the adult age (N=61). The development of surviving mice was strongly delayed. Eye opening did not occur before P25, compared to an average of P15 for wild-type littermates. Body growth was greatly reduced (Figure 3B). This correlated with delayed ossification of long bones (Figure 3C), and reduced growth hormone gene expression level in pituitary gland (Table I). After weaning, the body weight of the surviving mutant mice increased, but the adults suffered from permanent and disproportioned dwarfism. Measurement of body fat content by nuclear magnetic resonance did not reveal any increase in adiposity (Table I) also intraperitoneal content in white adipose tissues was found to be strongly reduced, when 4 weeks-old and 3 months-old animals were dissected. $TR\alpha^{AM1}xS$ females were sterile, without obvious histological defects in ovaries (data not shown).

Beside dwarfism, $TR\alpha^{AM1}xS$ animals displayed several other features of congenital hypothyroidism. This was the case in the spleen (Table I), whose weight was highly reduced at P15. In the cerebellum, the external granular layer, a transient structure which normally disappears at earlier stage, persisted at P21 (Figure 3D). The expression level of genes sensitive to hypothyroidism, *Hairless* in granular neurons and *Pcp2* in Purkinje cells was significantly reduced (Table I). Signs of cerebellar defect were also observed in adult $TR\alpha^{AM1}xS$ mice, which displayed a characteristic ataxia bearing, spreading their hindlimbs to maintain their posture. Adult $TR\alpha^{AM1}xS$ mice had reduced heart rate (Table I) and sometimes displayed cardiac arrhythmia. This cardiac phenotype correlated with a decreased expression at P15 for two T3 target genes, the potassium channel encoding genes *HCN2* and *KCNB1* (Table I). The body temperature was usually close to normal in $TR\alpha^{AM1}xS$ mice (Table I). However, a cold tolerance test revealed, like in mice lacking all TR isoforms (27) a defect in thermogenesis ability, the body temperature of $TR\alpha^{AM1}xS$ mice dropping within few hours after cold exposure for a fraction of animals (Figure 3E). T3 and T4 levels were not changed in $TR\alpha^{AM1}xS$ mice (Table I). The previously observed phenotypic alterations, which are all reminiscent of hypothyroidism, are thus not indirect consequences of a central deregulation of TH secretion but consequences of peripheral $TR\alpha1^{L400R}$ expression.

***TR β* expressing cells are less sensitive to *TR $\alpha1^{L400R}$* expression**

Previous genetic analysis underlined a predominant function of TR β 1 and TR β 2 receptors in several TH functions, including feedback-regulation of TSH secretion from pituitary thyrotropes and metabolic control in hepatocytes. TR β 1 is also involved in inner ear hair cells, retina cones and cerebellum Purkinje cells differentiation. Whether this results from predominant expression of *THRB*

over *THRA* in these cell types or from intrinsic properties of TR β receptors is unclear. Circulating level of TSH (data non shown) and *TSH β* expression in pituitary (Table I) were not significantly affected at P15 in *TRa^{AMI}xS* mice. In liver, Q-RT-PCR failed to demonstrate a significant change in expression level for two TH target genes at P15: *Dio1*, encoding type 1 deiodinase and *Me3*, encoding the NADP dependent malic enzyme (Table I). Similarly, Q-RT-PCR measurement of opsin gene expression failed to reveal a loss of M-Opsin cones in retina (data not shown), suggesting that, unlike *TR β* knockout mice, *TRa^{AMI}xS* are not colour blind. Purkinje cells differentiation was addressed by observing dendritic arborization after calbindin-D28k immunostaining and by measuring *Pcp2* mRNA level in cerebellum. These two parameters revealed a defect in differentiation at P15 (Figure 3F, table I). However, unlike what is reported for hypothyroid animals, this was followed by an apparent recovery at P21 (data not shown). TR β 1 function was also analyzed in the inner ear. Interestingly, it has been shown previously that, in outer hair cells, TR α 1 and TR β 1 can fulfill distinct functions. TR α 1 regulates *Kcnq4* a potassium channel, while TR β 1 activates *Slc26a5* expression, a gene encoding a motor protein called prestin (28). In line with this, immunocytochemistry revealed that the level of KCNQ4, but not of prestin, was highly reduced in *TRa^{AMI}xS* mice (figure 4). All these observations reveal that *TRa^{AMI}xS* mice do not display all the features of acute congenital hypothyroidism and that some TR β prevalent functions are preserved.

Tamoxifen induction of *TRa1^{L400R}* expression.

In order to evaluate the possibility to perform a kinetic study of TR α 1^{L400R} effect on development, we crossed *TRa^{AMI}/+* mice with *CagCreERTM* mice, that broadly express a modified CRE recombinase, which enzymatic activity is inducible by tamoxifen (29). The resulting heterozygous conditional mutants (*TRa^{AMI}xC*) were treated at E17.5 with tamoxifen. This resulted in the excision of the *PGKNeoRpolyA* cassette in most tissues. The excision efficiency varied from one tissue to the other (Figure 5A). In general the phenotype of tamoxifen treated *TRa^{AMI}xC* animals was similar to the one of *TRa^{AMI}xS*, but less severe. This conclusion was based on observations of growth curves (Figure 5B), ossification at P15 (Figure 5C), cerebellum histology (Figure 5D), adult heart rate, expression level at P15 of *KCNB1* and *HCN2* in heart, and *Hairless* and *Pcp2* in cerebellum (Table I). This demonstrates that expression of TR α 1^{L400R}, even when started at a late fetal stage, is sufficient to affect post-natal development. The relatively mild phenotype of tamoxifen treated *TRa^{AMI}xC* pups, compared to *TRa^{AMI}xS*, might only reflect the incomplete excision of the floxed cassette. We also performed tamoxifen treatment in adult *TRa^{AMI}xC* mice, to overcome detrimental effect of early TR α 1^{L400R} expression. This treatment did not have any visible effect in the short term cold tolerance assay, either because recombination efficiency was not sufficient in brown adipose tissue, or because the thermosensitive phenotype observed in *TRa^{AMI}xS* might result from a developmental alteration. Nevertheless, tamoxifen treatment of adult *TRa^{AMI}xC* males induced a significant reduction in heart

rate (Figure 3E) indicating that at least this last alteration is not a long term consequence of some developmental defect. *TRα^{AMI}_xC* mice therefore appear as a suitable model to separate between the developmental and adult homeostatic function of TRα.

DISCUSSION

In this report we describe for the first time a CRE inducible system able to express a dominant negative mutation of TRα1^{L400R}. TRα1^{L400R} is equivalent to an unliganded TRα1 receptor, as it prevents interactions with histone acetyl transferase transcription coactivators in a highly specific manner, without compromising interaction with corepressors. The first practical advantage of CRE dependent expression is to permit an easy production of mutant offspring, as the mutated receptor is not expressed in breeding animals. This also rules out that the maternal expression of the mutation indirectly influences the development of pups. This is expected for example for neuronal migration in the cortex (30). This experimental design also opens a broad field for new investigations of TH signaling in mice, as the use of tissue specific promoter and of an inducible version of the CRE will permit to control the expression of TRα1^{L400R} during development and in adults. This will permit to separate between the direct and indirect effects of TH signaling, as illustrated here for the cardiac function. As a large number of mice expressing CRE or an inducible version of CRE, have been already generated worldwide (31), this appears as a highly flexible method to create new animals models. These animals will be specifically affected in one or several of the functions sensitive to TRα1^{L400R} expression, within only one generation time. These functions include, in addition to those studied above, lipid storage in white adipose tissue (32), intestinal maturation (33), early post-natal erythropoiesis (9), lymphopoiesis (34) muscle function (35), myelin formation (36) and regeneration (37), hearing onset (38) , adult brain stem cells proliferation (39, 40), anxiety control and memory (41).

Three other *THRA* knock-in mutations have been reported before, which affect the TRα1 ligand-binding domain in different manner. Unlike the TRα1^{L400R} mutation, whose design was based on structural considerations, the changes in the TRα1 reading frame were copied from *THRB* germline mutations found in patients with resistance to thyroid hormone. Comparisons between *TRα^{AMI}_xS* and the previously reported *THRA* knock-in mouse strains enable to define constant features among the multiple consequences of expressing a dominant negative TRα1. As far as we can tell, the *TRα^{AMI}_xS* mice phenotype appears to be very similar to those reported for mice carrying the *TRα1^{R384C}*, which reduces the affinity of the ligand binding domain for T3 (15) and the *TRα1^{PV}* mutation, a frameshift mutation resulting in the loss of the N-terminal AF-2 domain (16, 42, 43). The fourth reported knock-in mutation, *TRα1^{P398H}*, is the only one that induces obesity and not dwarfism. From our data, we can rule out that this peculiar phenotype results from the inability of TRα1^{P398H} to recruit histone acetyl

transferase coactivators or release corepressors. It has been suggested that *TRα1*^{P398H/+} obesity rather results from a cross-talk between *TRα1*^{P398H} and *PPARα* in liver (44) (45). However a similar cross-talk with *PPARγ* in white adipocytes has been proposed to explain the opposite phenotype in *TRα1*^{PV/+} mice (42). Many human germline point mutations that prevent histone acetyl transferase recruitment have been reported for *THRB*, but not for *THRA*. From these studies we can predict that an AF-2 mutation in the human *TRα1* receptor would have dramatic consequences on development even at the heterozygous state.

The phenotypic similarities between *TRα*^{AMI}*xS* and *Pax8*^{-/-} hypothyroid mice demonstrate that the ligand-mediated recruitment of histone acetylase coactivators and/or release of corepressors by *TRα1* is crucial for post-natal development. This strengthens the previous conclusion that congenital hypothyroidism is mainly a manifestation of the negative action of unliganded *TRα1* (14). However differences are found between *TRα*^{AMI}*xS* and *Pax8*^{-/-} animals, suggesting a more complex situation. For example, the majority of *TRα*^{AMI}*xS* survive beyond weaning, while *Pax8*^{-/-} mice usually die within three weeks after birth. Our data suggest that in cells where *THRB* is highly expressed, like thyrotropes, hepatocytes, and retina cones, liganded *TRβ* can balance the down-regulation exerted by *TRα1*^{L400R}. Such a compensation cannot take place in hypothyroid mice, where unliganded *TRβ* receptors might instead participate to negative gene regulation and further compromise post-natal development. The presence of liganded *TRβ* in *TRα*^{AMI}*xS* can explain all the differences observed with *Pax8*^{-/-} hypothyroid mice, leaving little place for the proposed intervention of the antagonist *TRα2* isoform (19) or of an hypothetical *TR* independent *TH* signaling pathway (21, 46). It seems therefore that unliganded *TRα1*, and to a lesser extent unliganded *TRβ*, can account for all the detrimental effects of hypothyroidism in juveniles and in adults.

One important question that our data can help to clarify is whether the individual functions of *TRα1* and *TRβ1/2* isoforms in a given cell type are dictated by their respective abundance or by differences in their intrinsic properties. A vast amount of *in vitro* data suggests that *TRα1* and *TRβ1/2* are, at least at first sight, functionally equivalent. Knockout observations strengthen this hypothesis, as the phenotypic differences observed between *THRA* and *THRB* individual knockouts mainly reflect their contrasting expression patterns. Combinations of *THRA* and *THRB* knockout mutations also suggest functional redundancy, as several phenotypic alterations augmented (3, 47). As expected *TRα1*^{L400R} exerts in transient expression assays a dominant-negative action both on *TRα1* and *TRβ1*. Accordingly, most features of the *TRα*^{AMI}*xS* phenotype can be predicted from the respective abundance of *THRA* and *THRB* encoded receptors in a given cell type. First, we failed to detect any phenotypic alteration in cell types known to express *THRB* at much higher level than *THRA*. This includes retina cones, hepatocytes and thyrotrope cells. Second, *TRβ* receptors appear sensitive to the *TRα1*^{L400R}

dominant-negative effect in tissues where the stoichiometry is less favorable to *THRB*. Previous genetic studies have shown that compound knockout mice devoid of all receptors have a reduced body temperature and are much more sensitive to cold exposure than *THRA* knockout mice (27, 48, 49). Thus, despite pharmacological evidences indicating only partial overlap (50), the functions of the two receptors seem to be redundant for cold resistance, correlating with the concomitant presence of TR α 1 and TR β 1 in brown adipose tissue (51). The fact that TR α ^{AMI}*xS* are highly sensitive to cold exposure therefore suggests that TR α 1^{L400R} can interfere with both TR α 1 and TR β 1 functions in brown adipocytes. A similar interpretation can be proposed for Purkinje cells. Hypothyroidism results in a permanent reduction in dendritic arborization and a disorganization of the cells alignment (52). *In situ* hybridization reveals a predominant *THRB* expression in these cells after birth (53), and their differentiation is deeply affected by a TR β knock-in mutation (20). However, *in vitro* differentiation of purified Purkinje precursors rather suggests a predominant function for TR α 1 over TR β 1 (54). We show here that TR α 1^{L400R}, like TR α 1^{R384C} (41) has only a transient effect on Purkinje dendrites arborization. A possible explanation would be that, as *THRB* expression increases over time in this cell type (55), TR β 1 progressively accumulates, overcomes the transcription repression exerted by TR α 1^{L400R} and eventually unlocks differentiation. In conclusion, expression patterns clearly influence the respective *in vivo* function of TR α 1 and TR β 1/2. TR stoichiometry is however unlikely to be the only explanation for the maintenance of several TR β 1/2 dependent functions in TR α ^{AMI}*xS* mice. Some our results also suggest that TR α 1 and TR β 1/2 intrinsic properties, and the set of target genes that they control, are different. For example, the resting body temperature of most TR α ^{AMI}*xS* animals is normal, unlike what is reported for compound *THRA/THRB* knockout mice, suggesting that part of the activation by TR β 1 is preserved in brown adipocytes. Most importantly, the inner ear phenotype provides compelling evidence that TR α 1 and TR β 1 intrinsic properties are different. Within outer hair cells, TR α 1^{L400R} appears to antagonize TR α 1 driven transactivation of KCNQ4 without compromising TR β 1 mediated regulation of prestin. It seems therefore that in vertebrates, divergent evolution of the paralogous *THRA* and *THRB* genes increased the variety of cellular responses to TH.

MATERIAL AND METHODS

Plasmids and transient expression assays

The L400R mutation was introduced in the TR α 1 reading frame present in the pBIRDTR α 1 construct (56) using for PCR mutagenesis an oligonucleotide containing a SacII restriction site at the mutation site: 5' CC CCG CGG TTC CTG GAG GTC TTT GAG 3'. The entire cDNA structure was confirmed by DNA sequencing. The mutated cDNA was inserted between a floxed *PGKNeoRpolyA* (57) able to stop transcription, and an *IRESTaulacZ* sequence (58) to create a bicistronic mRNA encoding both the mutated receptor TR α 1^{L400R} and the reporter Tau- β -galactosidase fusion protein targeted to the

cytoskeleton. The completed construct was transferred as a PacI restriction fragment into the pBK-CMV expression vector (Stratagene San Diego) (figure 1B) or between two large genomic fragments (Figure 2A) cloned into the Supercos1 cosmid vector (Stratagene) for homologous recombination. An identical pBK-CMV derivative was made for the intact TR α 1 cDNA for control experiments. For transient expression, Cos-7 cells, maintained in TH depleted medium, were transfected with Exgen reagent (Fermentas). pTK-DR4(2x)-luc was used as a reporter construct (59) and pRL-CMV (Promega), encoding renilla luciferase, as an internal standard. T3 (10^{-7} M) was eventually added to the medium 24h before the quantification of luciferase activity (Promega, Dual Luciferase Assay). The full-length TR α 1 and TR α 1^{L400R} reading frames were transferred into pAS2.1 (Clontech) to perform two-hybrid assays in yeast cells. Two-hybrid interactions were tested by plating AH109 yeast cells on selective medium and quantified by measuring β -galactosidase activity in Y187 yeast cells, using orthonitrophenyl- β -D-galactopyranoside as a substrate.

Generation of mutant mice

The oligonucleotides used for vector construct, screening and TR α expression analysis are the following (positions on figure 2). a: 5'GCGATACCGTAAAGCACGAG ; b: 5'GCCTTCTATCGCCTTCTTGACG ; c: 5'CGTCTGGAGAAGAGTTGG d: 5'TCCACAGGTATCTCCAGACAGG e: 5' GATTCTTCTGGATTGTGCGGCG f: ACCGCAAACACAACATTCGCACTTCTGGC g: 5' GAGGAAGGAGAGAAGAGATG GGGGTTC h: 5' CGGTCGCTACCATTACC AGTTG i: 5' TTATGGATGGACGGACGG j: 5' AGTCTACGGCAAGGCAACACCAAG. 5×10^6 129/Sv mouse embryonic stem cells (13) were electroporated with 40 μ g of linear plasmid and selected with G418 (250 μ g/ml; Gibco-BRL). Cell clones were picked 10 days later and screened by PCR amplification of junction fragment, using Long expand Taq polymerase (Roche) (a+c for the 5' side, h+j for the 3' side). 3 out of 310 clones carried the mutant allele called TR α ^{AMI} (*AF-2 Mutation, Inducible*). After further PCR characterization of the recombinant allele, cells were injected into C57/B16 blastocysts to generate chimeras. After germline transmission, transgenic mice were routinely screened by PCR directed on the lacZ – exon 9 junction (h+i). TR α ^{AMI}_{xS} and TR α ^{AMI}_{xC} mice were produced by crossing TR α ^{AMI}/+ mice with SYCP1CRE transgenic mice (26) and CagCreERTM (29) respectively and identified by PCR (5'TTACCGGTCGATGCAACG3'+ 5'CCAGCCACCAGCTTGCAT3' for CRE). When indicated, TR α ^{AMI}/C mice received 1 (pregnant E17 mothers) or 5 (adults) daily intra-peritoneal injection of tamoxifen (Sigma T-5648, 50 mg/kg) dissolved in corn oil. CRE mediated deletion was identified by PCR (b+d+e). All animals experimentations were performed under Animal care procedures and conducted in accordance with the guidelines set by the European Community Council Directives (86/609/EEC).

Phenotype analysis

RNA were extracted from tissues of 8-10 mice per group and purified using RNeasy (Qiagen) or RNA Nanoprep (Stratagene) extraction kits, including a DNaseI treatment. RNA quality controls were performed by gel electrophoresis (Agilent 2100 Bioanalyzer). cDNA were prepared from 1 µg RNA using M-MLV reverse transcriptase (Promega) and random 6-mers primers. After 1/40 dilution, 2 µl of cDNA were used for quantitative PCR (Stratagene Mx3000P® QPCR System), using either Platinum® Quantitative PCR SuperMix (Invitrogen) or Taqman Assay-on-Demand (Applied Biosystems). Quantitation was performed in duplicates using the *HPRT* and *TBP* housekeeping genes as internal standards (or *ARBP* for pituitary) and the $2^{-\Delta\Delta Ct}$ method for data analysis (60). Paraffin sections were prepared from tissues fixed with 4% paraformaldehyde. Purkinje cells were stained with a rabbit anti-calbindin-D-28k antibody (Swant CB38a, 1/5000 dilution). Serum T4, T3 and TSH levels were measured as described previously (61). Cold tolerance assays were performed with a telemetry system as previously described (48) in a 4°C room. Body temperature was measured every 15 or 30 minutes during 8 hours and mice were removed from the cold room when their body temperature dropped below 35°C. Inner ear immunocytochemistry was performed as described previously (28).

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FIGURES

Figure 1: Properties of the TR α 1^{L400R} mutated receptor

A: Two-hybrid interactions in yeast show that TR α 1^{L400R} can only interact with corepressors. Unlike liganded TR α 1, TR α 1^{L400R} does not interact with SRC1 and SRC2 histone acetyl transferase coactivators. The interaction with the NcoR corepressor is ligand sensitive for TR α 1 but constitutive for TR α 1^{L400R}. B: Cytomegalovirus (CMV) and phospho-glycerate-kinase (PGK) promoters generate transcripts terminated by the SV40 polyadenylation signal present in the *PGKNeoRpolyA* cassette. The bicistronic TR α 1^{L400R} and TaulacZ cDNA, located downstream to the SV40 polyadenylation signal, cannot be expressed. As the *PGKNeoRpolyA* cassette is flanked by two loxP sequences, CRE mediated deletion enables TR α 1^{L400R} and TaulacZ synthesis. C: TR α 1^{L400R} expression requires CRE mediated deletion. Western blotting of mammalian COS7 cells transfected with the CMV construct encoding for TR α 1^{L400R} (1, 2) or the empty CMV construct (3). When the floxed *PGKNeoRpolyA* cassette is present (1), only a non-specific band is detected, reflecting the absence of TR α 1^{L400R} expression. The CRE deleted construct (2) encodes the expected 44 kDa TR α 1^{L400R} protein. D: TR α 1^{L400R} acts as a dominant negative receptor. Transient luciferase expression in transfected COS7 cells show that, unlike wild-type TR α 1, TR α 1^{L400R} can not transactivate a DR4-TK-luc construct in the presence of T3. TR α 1 mediated transactivation is antagonized by increasing amount of TR α 1^{L400R}. A 1/1 ratio leads to a complete inhibition of T3 response.

Figure 2: Generation of the TR α ^{AMI} allele by homologous recombination in mouse embryonic stem cells.

A: Structure of the *THRA* locus after homologous recombination. From 5' to 3': *THRA* sequences absent from the recombination vector (black line) 6,5kb of *THRA* genomic sequences extending to the non coding part of exon2 (nucleotide 279 on Genebank # NC_000077 dark grey box); the *PGKNeoRpolyA* cassette, providing G418 resistance to ES cells, flanked by two tandem loxP for CRE mediated deletion; TR α 1^{L400R} encoding sequence; *ECMV-IRES* element for internal ribosomal entry; *TaulacZ* coding sequence; 2.9 kb of *THRA*, starting in exon 9 (nucleotide 22889 on Genebank # NC_000077 dark grey box), 3' genomic sequences absent from the recombination vector (black line). White arrows indicate the position and orientation of oligonucleotides used for PCR amplification. The SacII restriction site covers the L400R encoding codon. CRE mediated recombination results in the deletion of the floxed selection cassette, allowing for transcription and translation of the downstream TR α 1^{L400R} and TaulacZ reporter protein. B PCR confirmation of homologous recombination in the *THRA* locus in ES cells. Primers a and c amplify a 10.8 kb fragment (2) while h and j amplify 3.3 kb (4) after homologous recombination but not after random DNA integration (1 and 3). C CRE mediated recombination in TR α ^{AMI/+} ES cells triggers TR α 1^{L400R} expression. RT-PCR of RNA purified from ES cells (primers f and g) amplifies 274 bp of TR α 1 cDNA (*) and a non specific product (***)). SacII digestion releases two fragments (135bp+139bp **) in ES cells carrying one

CRE-deleted $TR\alpha^{AMI}$ allele (3). $TR\alpha^{L400R}$ expression is not detected in ES cells carrying wild-type alleles (1) or one intact $TR\alpha^{AMI}$ allele (2).

Figure 3: phenotype of $TR\alpha^{AMI}xS$ mice

A $TR\alpha^{AMI}xS$ mice have lost the *PGKNeopolyA* cassette. PCR was performed on tail DNA with primers b,d,e (Figure 2) amplifying either 1200bp (b+e) for the full length $TR\alpha^{AMI}$ allele or 800bp (d+e) for the CRE deleted $TR\alpha^{AMI}$ allele. Due to the presence of intronic sequences, the intact *THRA* locus can not be amplified in these conditions. B Body growth reduction in $TR\alpha^{AMI}xS$ mice compared to wild-type littermates. Body weight (g) was measured during 80 days (N=22). C Alizarine staining, performed on whole mount skeleton, reveals delayed ossification of long bones (arrow) in $TR\alpha^{AMI}xS$ mice at P15, clearly visible in posterior limbs. D Delayed cerebellum development in $TR\alpha^{AMI}xS$ mice: sagittal section shows the persistence of the external granular layer (arrows) at P21 in $TR\alpha^{AMI}xS$ cerebellum (haematoxylin staining) but not in control littermates. E Cold tolerance test: The body temperature of 5 $TR\alpha^{AMI}xS$ 5-months old adult females (black lines) and 4 control female littermate (grey lines) was recorded during 8 hours of cold exposure. 3 out of 5 animals failed to maintain their body temperature within this time period. F Purkinje cells dendritic arborization, revealed by calbindin-D28k staining, is visible above cell nuclei. The number and size of dendritic spines is reduced at P15 in $TR\alpha^{AMI}xS$ cerebellum.

Figure 4: Inner ear phenotype of $TR\alpha^{AMI}xS$ mice

Immunohistochemistry reveals the presence of both prestin and KCNQ4 (red) in wild-type outer hair cells (OHC) at P15. The same observation was performed on 3 $TR\alpha^{AMI}xS$ mice and 4 wild-type mice. Small arrows underline basolateral staining for prestin, and perinuclear staining for KCNQ4. Vertical arrows indicate the positions of cells nuclei, counterstained with 4',6-diamidino-2-phenylindole (DAPI, blue). Whereas prestin staining is maintained, perinuclear KCNQ4 staining is lost in $TR\alpha^{AMI}xS$ mice (*).

Figure 5: phenotype of $TR\alpha^{AMI}xC$ mice

A-D: Phenotype of $TR\alpha^{AMI}xC$ pups born from females treated with tamoxifen at gestational day 17.5. A: Deletion of the *PGKNeoRpolyA* cassette after tamoxifen treatment at E17.5, analyzed by PCR at P15 (primers b, d, e Figure 2) reveals the presence of a 800bp fragment (*) corresponding to the CRE deleted $TR\alpha^{AMI}$ allele in cerebellum (cb) whole brain (wb) and small intestine (si) in $TR\alpha^{AMI}xC$ mice (+), but not in control littermates without CRE transgene (-). The upper 1200bp band corresponds to the full-length $TR\alpha^{AMI}$ allele. B Body weight (g) was measured during 138 days (N=10). Body growth is less affected in $TR\alpha^{AMI}xC$ mice than in $TR\alpha^{AMI}xS$ mice (Figure 3B) but significantly reduced compared to $TR\alpha^{AMI}/+$ littermates, which do not express the mutation. C Alizarine staining performed on whole mount skeleton reveals delayed ossification of long bones in $TR\alpha^{AMI}xC$ mice at P15. The

defect, marked by arrows, is less pronounced than for $TR\alpha^{AMI}xS$ mice (Figure 3C). D Persistence of external granular layer in cerebellum at P21. Unlike what is found in wild-type controls, granular precursor cells (black arrow, here in the groove between lobe IV and lobe V) are still present in $TR\alpha^{AMI}xC$, but less abundant than in $TR\alpha^{AMI}xS$. E: Tamoxifen treatment of 8 weeks-old adult males induces a significant decrease ($p=0.01$ Student t test) of heart rate, measured one week later, in $TR\alpha^{AMI}xC$ (N=7) compared to age matched wild-type controls (N=7), also treated with tamoxifen. $TR\alpha^{AMI}xS$ data (N=4) are given for comparison.

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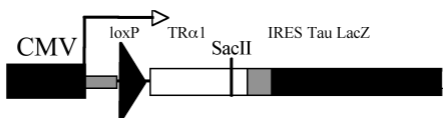
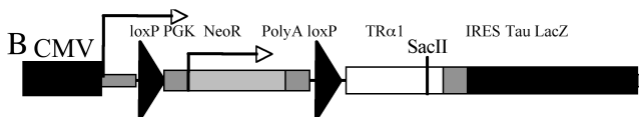
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Mouse genotype	+/+	<i>TRα^{AM1}/+</i>	<i>TRα^{AM1}xS</i>	<i>TRα^{AM1}xC*</i>	
Physiological parameters P15					
Serum T4 (µg/dL)	9,3±0,6	9,6±1	9,0±2,4	6,8±2,2**	
Serum T3 (ng/dL)	141±22	168±15	129±13	164±8	
Spleen weight (mg)	39,5±9,1	45,0±10,8	26,9±12,2****	26,2±7,5**	
Physiological parameters in adult males (2 – 3 months)					
Serum T4 (µg/dL)	4,3±0,8	ND	4,2±0,7	ND	
Serum T3 (ng/dL)	116±24	ND	123±12	ND	
Serum TSH (mU/L) †	144±106	ND	50±26	ND	
Heart beat (/min) †	393±45	ND	224±24****	ND	
Body temperature (°C)	38,3±0,4	38,6±0,2	38,5±0,4	38,4±0,5	
Body fat content (%)†	9,33±1,61	ND	8,20±0,36	ND	
Gene expression P15 (% of wild type)					
Gene	Tissue/organ				
<i>GH</i>	pituitary	100±47	142±33	44±20**	52±18
<i>TSHβ</i>	pituitary	100±31	171±53**	65±32	120±12
<i>Dio1</i>	liver	100±17	112±26	101±51	ND
<i>ME3</i>	Liver	100±38	86±17	130±20	ND
<i>TRα1+TRα1^{L400R}</i>	liver	100±12	71±15**	94±6	81±13
<i>HCN2</i>	heart	100±27	76±32	7±2***	5±2***
<i>KCNB1</i>	heart	100±18	87±24	25±6***	21±7***
<i>Hr</i>	cerebellum	100±19	132±37	19±5***	39±11***
<i>PcP2</i>	cerebellum	100±21	122±21	38±21***	105±10
<i>TRα1+TRα1^{L400R}</i>	cerebellum	100±26	140±29**	327±48***	374±56***

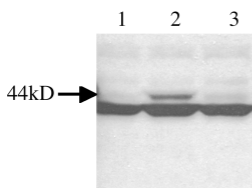
* tamoxifen treated at E17.5 ** different from wild type, $p < 0,05$ *** $p < 0,01$ ND = Not Determined †Males only

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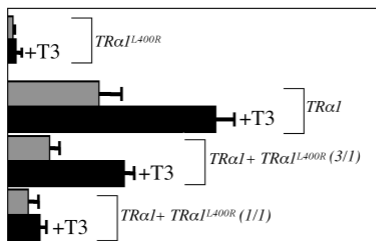
	SRC1		SRC2		NcoR	
T3	-	+	-	+	-	+
TR α 1	-	+	-	+	+	-
TR α 1 ^{L400R}	-	-	-	-	+	+

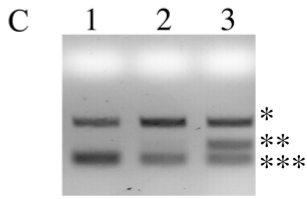
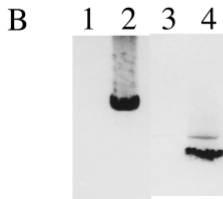
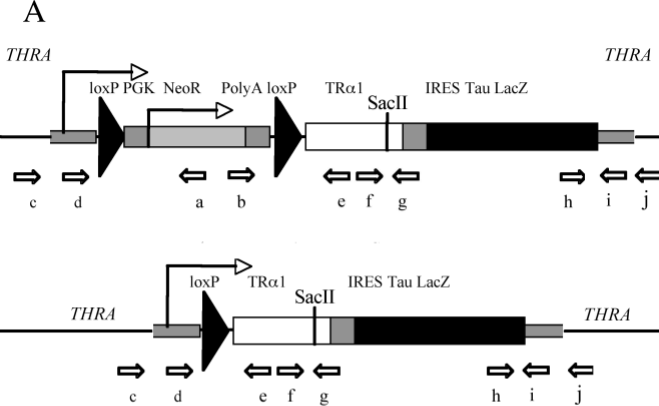


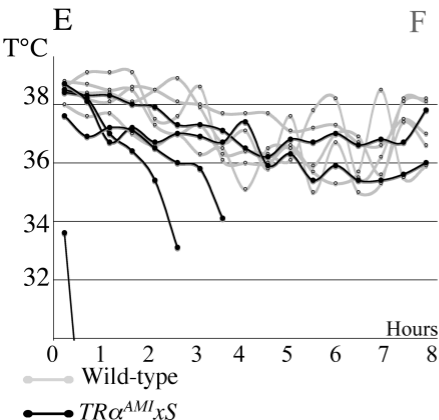
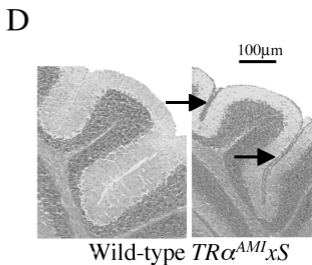
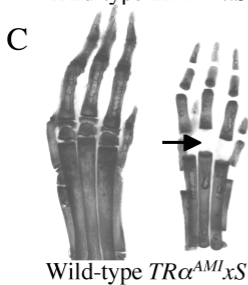
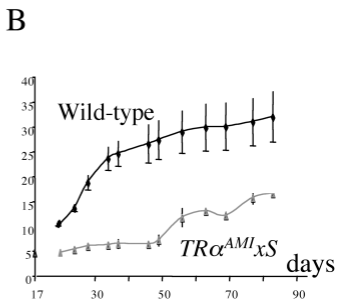
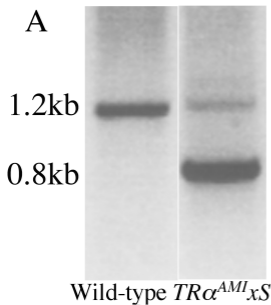
C



D



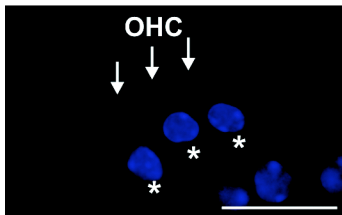
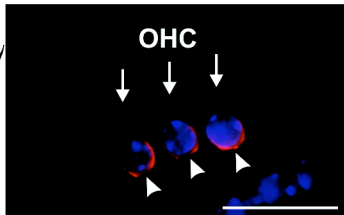




Wild type

$TR\alpha^{AMI}$

KCNQ4



Prestin

